



# IG® Autoinduction DE3 ElectroCompetent Cells

## Manual

|                     |                |                |
|---------------------|----------------|----------------|
| <b>Catalog #</b>    | <b>1265-12</b> | <b>1265-48</b> |
| <b>Package Size</b> | 6x50 µl        | 24x50 µl       |



**Important!**

**-80°C Storage Required**

- \* Immediately inspect packages
- \* Freeze upon receipt



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## Description:

Intact Genomics (IG®) ElectroCompetent Autoinduction DE3 cells are suitable for transformation and routine recombinant protein expressed from a vector under an IPTG-inducible T7 promoter. This Autoinduction strain with the DE3 lysogen, which contains a chromosomally integrated IPTG-inducible promoter enabling T7 RNA polymerase, is expressed by IPTG induction, as well as autoinduction with IG® optimized Autoinduction media. (catalog #: 1722). With simple and less hands-on autoinduction, you can either enjoy more free-time or get more work done. Moreover, the yields of both expressed cells and protein are likely 5-10 fold or more for a broad range of protein targets at either small or large scale of protein production.

## Specifications:

**Competent cell type:** ElectroCompetent

**Strain:** IG Autoinduction BL21(DE3)

**Species:** *E. coli*

**Format:** Tubes

**Transformation efficiency:**  $\geq 1.0 \times 10^{10}$  cfu/ $\mu$ g pUC19 DNA

**Blue/white screening:** Yes

**Shipping condition:** Dry ice

## Reagents Needed for One Reaction:

- IG Autoinduction DE3 ElectroCompetent cells: 25  $\mu$ l
- DNA (or pUC19 Control, 10 pg/ $\mu$ l): 1  $\mu$ l
- Recovery medium: 1 ml

## Product Components & Storage:

- IG Autoinduction DE3 ElectroCompetent cells: -80 °C
- pUC19 control DNA: -20 °C
- Recovery medium: 4 °C

## Genomic Features:

IG Autoinduction DE3 ElectroCompetent cells have the following features:

- T7 Expression Strain
- Optimized for autoinduction
- Suitable for expression of toxic genes
- TetR: Tetracycline resistant (12.5  $\mu$ g/ml)

## Quality Control:

Transformation efficiency is tested by using the pUC19 control DNA supplied with the kit and the high efficiency transformation protocol listed below. Transformation efficiency should be  $\geq 1 \times 10^{10}$  CFU/ $\mu$ g pUC19 DNA.

Untransformed cells are tested for appropriate antibiotic sensitivity.

## General Guidelines:

Follow these guidelines when using IG Autoinduction DE3 ElectroCompetent *cells*:

- Handle competent cells gently as they are highly sensitive to changes in temperature or mechanical lysis caused by pipetting.
- Thaw competent cells on ice and transform cells immediately following thawing. After adding DNA, mix by tapping the tube gently. Do not mix cells by pipetting or vortexing.

**Note:** A high-voltage electroporation apparatus such as Bio-Rad Gene Pulser II #165-2105, capable of generating field strengths of 16 kV/cm is required.

## Transformation Protocol:

Use this procedure to transform IG Autoinduction DE3 electrocompetent cells. Do not use these cells for chemically transformation.

- 1) Place sterile cuvettes and microcentrifuge tubes on ice.
- 2) Remove competent cells from the -80 °C freezer and thaw completely on wet ice (10-15 minutes).
- 3) Aliquot 1  $\mu$ l (1 pg-10 ng) of DNA to the chilled microcentrifuge tubes on ice.
- 4) When the cells are thawed, add 25  $\mu$ l of cells to each DNA tube on ice and mix gently by tapping 4-5 times. For the pUC19 control, add 1  $\mu$ l of (10 pg/ $\mu$ l) DNA to the 25  $\mu$ l of cells on ice. Mix well by tapping. Do not pipette up and down or vortex to mix, this can harm cells and decrease transformation efficiency.
- 5) Pipette 26  $\mu$ l of the cell/DNA mixture into a chilled electroporation cuvette without introducing bubbles. Quickly flick the cuvette downward with your wrist to deposit the cells across the bottom of the well and then electroporate.
- 6) Immediately add 974  $\mu$ l of Recovery Medium or any other medium of choice to the cuvette, pipette up and down three times to re-suspend the cells. Transfer the cells and Recovery Medium to a culture tube.
- 7) Incubate tubes at 37 °C for 1 hour at 210 rpm.
- 8) Dilute the cells as appropriate then spread 20-200  $\mu$ l cells onto a pre-warmed selective plate. For the pUC19 control, plate 50  $\mu$ l of diluted transformants onto an LB plate containing 100  $\mu$ g/ml ampicillin. Use sterilized spreader or autoclaved ColiRoller™ plating beads to spread evenly.
- 9) Incubate the plates overnight at 37 °C.

## Calculation of Transformation Efficiency:

Transformation Efficiency (TE) is defined as the number of colony forming units (cfu) produced by transforming 1 µg of plasmid into a given volume of competent cells.

$$TE = \text{Colonies}/\mu\text{g}/\text{Dilution}$$

Transform 1 µl of (10 pg/µl) pUC19 control plasmid into 50 µl of cells, add 950 µl of Recovery Medium. Dilute 10 µl of this in 990 µl of Recovery Medium and plate 50 µl. Count the colonies on the plate the next day. If you count 100 colonies, the TE is calculated as follows:

$$\text{Colonies} = 100$$

$$\mu\text{g of DNA} = 0.00001$$

$$\text{Dilution} = 50/1000 \times 10/1000 = 0.0005$$

$$TE = 100/.00001/.0005 = 2.0 \times 10^{10}$$

## Related Products:

- ig® 5-Alpha Chemically Comp. Cells (Cat.# 1031-12)
- BL21(DE3) Electroporation Comp. Cells (Cat.# 1212-12)
- T4 DNA Ligase (Cat.# 3212)
- i7® High Fidelity DNA Polymerase (Cat.# 3254)
- IG® Autoinduction Chemically Competent Cells (Cat.#: 1065)
- IG® Autoinduction Media (Cat.#: 1722)
- igFusion™ Cloning Kit (Cat.# 4111)

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Our hours are Monday - Friday, 8AM to 5PM, U.S. Central Standard Time.

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