



ig[®] Stable 2 Electrocompetent *E. coli* Cells

Manual

Catalog #	1216-12	1216-24
Package Size	6x50μl	12x50μl



Important!

-80°C Storage Required

- * Immediately inspect packages
- * Freeze upon receipt

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Description:

Intact Genomics (ig $^{\circ}$) Stable 2 electrocompetent *E. coli* cells offer the highest transformation efficiencies of $\geq 2x \ 10^{10}$ cfu/µg plasmid DNA which are ideal for applications requiring high transformation efficiencies, such as with cDNA or gDNA library construction. Stable 2 cells are capable of cloning methylated genomic sequences, retroviral sequences and direct repeat sequences. Intact Genomics Stable 2 cells provide superb transformation efficiency, allowing for increased opportunity for experimental success.

Specifications:

Competent cell type: Electrocompetent

Species: E. coli

Format: Tubes

Transformation efficiency: ≥2 x 10¹⁰ cfu/μg pUC19 DNA

Blue/white screening: No

Shipping condition: Dry ice

Reagents Needed for One Reaction:

• ig[®] Stable 2 electrocompetent cells: 25 μl

• DNA (or pUC19 Control, 10 pg/μl): 1 μl

Recovery medium: 1 ml

Product Components & Storage:

ig® Stable 2 electrocompetent cells: -80 ºC

pUC19 control DNA: -20 ºC

Recovery medium: 4 ºC

Genotype:

F- mcrA Δ(mcrBC-hsdRMS-mrr) recA1 endA1lon gyrA96 thi supE44 relA1 λ- Δ(lac-proAB)

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Product Benefits:

ig® Stable 2 electrocompetent cells have the following features:

- Stable 2 allows for cloning of methylated genomic sequences
- Stabilizes retroviral and direct repeat sequences including HIV
- High transformation efficiency allows aids in cloning rare sequences
- May be used for plasmids > 20 kb
- endA1 mutation increases plasmid yield significantly

Quality Control:

Transformation efficiency is tested by using the pUC19 control DNA supplied with the kit and the high efficiency transformation protocol listed below. Transformation efficiency should be $\geq 2 \times 10^{10}$ CFU/µg pUC19 DNA. Untransformed cells are tested for appropriate antibiotic sensitivity.

General Guidelines:

Follow these guidelines when using ig Stable 2 ElectroCompetent Cells:

- Handle competent cells gently as they are highly sensitive to changes in temperature or mechanical lysis caused by pipetting.
- Thaw competent cells on ice, and transform cells immediately following thawing. After adding DNA, mix by tapping the tube gently. Do not mix cells by pipetting or vortexing.

Note: A high-voltage electroporation apparatus such as Bio-Rad Gene Pulser II #165-2105, capable of generating field strengths of 16 kV/cm is required.



Transformation Protocol:

Use this procedure to transform ig® Stable 2 electrocompetent cells. Do not use these cells for chemical transformation.

- 1) Place sterile cuvettes and microcentrifuge tubes on ice.
- 2) Remove competent cells from the -80 °C freezer and thaw completely on wet ice (10-15 minutes).
- 3) Aliquot 1 μ l (1 pg-10 ng) of DNA to the chilled microcentrifuge tubes on ice.
- 4) When the cells are thawed, add 25 μ l of cells to each DNA tube on ice and mix gently by tapping 4-5 times. For the pUC19 control, add 1 μ l of (10 pg/ μ l) DNA to the 25 μ l of cells on ice. Mix well by tapping. Do not pipette up and down or vortex to mix, this can harm cells and decrease transformation efficiency.
- 5) Pipette 26 μ l of the cell/DNA mixture into a chilled electroporation cuvette without introducing bubbles. Quickly flick the cuvette downward with your wrist to deposit the cells across the bottom of the well and then electroporate.
- 6) Immediately add 974 μ l of Recovery Medium or any other medium of choice to the cuvette, pipette up and down three times to re-suspend the cells. Transfer the cells and Recovery Medium to a culture tube.
- 7) Incubate tubes at 37 °C for 1 hour at 210 rpm.
- 8) Dilute the cells as appropriate then spread 20-200 μl cells onto a pre-warmed selective plate. For the pUC19 control, plate 50 μl of diluted transformants onto an LB plate containing 100 μg/ml ampicillin. Use sterilized spreader or autoclaved ColiRoller[™] plating beads to spread evenly.
- 9) Incubate the plates overnight at 37 °C.



Example Calculation of TE:

Transformation Efficiency (TE) is defined as the number of colony forming units (cfu) produced by transforming 1µg of plasmid into a given volume of competent cells.

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TE = Colonies/μg/Dilution
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Transform 1 μ l of (10 pg/ μ l) pUC19 control plasmid into 25 μ l of cells, add 950 μ l of Recovery Medium. Dilute 10 μ l of this in 990 μ l of Recovery Medium and plate 50 μ l. Count the colonies on the plate the next day. If you count 100 colonies, the TE is calculated as follows:

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Colonies = 100

\mug of DNA = 0.00001

Dilution = 50/1000 x 10/1000 = 0.0005

TE = 100/.00001/.0005 = 2.0x10<sup>10</sup>
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Related Products:

- ig® 5-Alpha Chemically Comp. Cells (Cat.# 1031-12)
- ig® 10B Chemically Comp. Cells (Cat.# 1011-12)
- T4 DNA Ligase (Cat.# 3212)
- i7[®] High Fidelity DNA Polymerase (Cat.# 3254)
- Quick10™ Cloning Kit (Cat.# 4122)

Ordering Information:

- Order online within the USA. Place orders on www.intactgenomics.com using our secure Shopping Cart.
- Order by email, phone, or fax.

Email: sales@intactgenomics.com

Phone: (314) 942-3655 | Toll-free: 855-835-7172 | Fax: (314) 942-3656

Order via our distributors.

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