ig® XL1 Blue Max Electrocompetent *E. coli* Cells

Manual

Catalog #	1223-12	1223-24
Package Size	6x50µl	12x50µl

Important!

-80°C Storage Required

* Immediately inspect packages
* Freeze upon receipt

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Intact Genomics, Inc.

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Description:

Intact Genomics (ig®) XL1 Blue Max electrocompetent *E. coli* cells offer the highest transformation efficiencies of ≥5 x 10^{10} cfu/µg plasmid DNA. These are ideal for applications requiring high transformation efficiencies such as cDNA or gDNA library construction. These cells have the capability to allow for the preparation of high quality plasmid DNA, and single strand rescue of phagemid DNA and Blue/white screening. XL1 Blue Max cells provide transformation efficiencies that are significantly higher than any competitor's similar product, allowing for increased opportunity for experimental success.

Specifications:

Competent cell type: Electrocompetent

Species: *E. coli*

Format: Tubes

Transformation efficiency: ≥ 2.5 x 10^{10} cfu/µg pUC19 DNA

Blue/white screening: Yes

Shipping condition: Dry ice

Reagents Needed for One Reaction:

- ig® XL1 Blue Max Electrocompetent cells: 25 µl
- DNA (or pUC19 Control, 10 pg/µl): 1 µl
- Recovery medium: 1 ml

Product Components & Storage:

- ig® XL1 Blue Max Electrocompetent cells: -80 ºC
- pUC19 control DNA: -20 ºC
- Recovery medium: 4 ºC

Genotype:

recA1 endA1 gyrA96 thi-1 hsdR17 supE44 relA1 lac [F´ proAB lacIq ZΔM15 Tn10 (Tetr)]

Product Benefits:

ig® XL1 Blue Max electrocompetent cells have the following features:

- XL1 Blue Max cells are tetracycline resistant.
- XL1 Blue Max cells are endonuclease (endA) deficient, which greatly improves the quality of miniprep DNA.
- XL1 Blue Max cells recombination (recA) deficient, improving insert stability.
- Cleavage of cloned DNA by the EcoK endonuclease system is prevented by the hsdR mutation.
- Blue-white color screening via the lacIq ZΔM15 gene on the F´ episome.

Quality Control:

Transformation efficiency is tested by using the pUC19 control DNA supplied with the kit and using the protocol given below. Transformation efficiency should be ≥5 x 10^{10} CFU/µg pUC19 DNA. Untransformed cells are tested for appropriate antibiotic sensitivity.

General Guidelines:

Follow these guidelines when using ig® XL1 Blue Max Electrocompetent Cells:

- Handle competent cells gently as they are highly sensitive to changes in temperature or mechanical lysis caused by pipetting.
- Thaw competent cells on ice and transform cells immediately following thawing. After adding DNA, mix by tapping the tube gently. Do not mix cells by pipetting or vortexing.

Note: A high-voltage electroporation apparatus such as Bio-Rad Gene Pulser II #165-2105, capable of generating field strengths of 16 kV/cm is required.

Transformation Protocol:

Use this procedure to transform ig® XL1 Blue Max electrocompetent cells. Do not use these cells for chemical transformation.

1. Place sterile cuvettes and microcentrifuge tubes on ice.
2. Remove competent cells from the -80 °C freezer and thaw completely on wet ice (10-15 minutes).
3. Aliquot 1 µl (1 pg-10 ng) of DNA to the chilled microcentrifuge tubes on ice.
4. When the cells are thawed, add 25 µl of cells to each DNA tube on ice and mix gently by tapping 4-5 times. For the pUC19 control, add 1 µl of (10 pg/µl) DNA to the 25 µl of cells on ice. Mix well by tapping. Do not pipette up and down or vortex to mix, this can harm cells and decrease transformation efficiency.
5. Pipette 26 µl of the cell/DNA mixture into a chilled electroporation cuvette without introducing bubbles. Quickly flick the cuvette downward with your wrist to deposit the cells across the bottom of the well and then electroporate.
6. Immediately add 974 µl of Recovery Medium or any other medium of choice to the cuvette, pipette up and down three times to re-suspend the cells. Transfer the cells and Recovery Medium to a culture tube.
7. Incubate tubes at 37 °C for 1 hour at 210 rpm.
8. Dilute the cells as appropriate then spread 20-200 µl cells onto a pre-warmed selective plate. For the pUC19 control, plate 50 µl of diluted transformants onto an LB plate containing 100 µg/ml ampicillin. Use sterilized spreader or autoclaved ColiRoller™ plating beads to spread evenly.
9. Incubate the plates overnight at 37 °C.

Example Calculation of TE:

Transformation Efficiency (TE) is defined as the number of colony forming units (cfu) produced by transforming 1µg of plasmid into a given volume of competent cells.

TE = Colonies/µg/Dilution

Transform 1 µl of (10 pg/µl) pUC19 control plasmid into 25 µl of cells, add 950 µl of Recovery Medium. Dilute 10 µl of this in 990 µl of Recovery Medium and plate 50 µl. Count the colonies on the plate the next day. If you count 100 colonies, the TE is calculated as follows:

Colonies = 100

µg of DNA = 0.00001

Dilution = 50/1000 x 10/1000 = 0.0005

TE = 100/.00001/.0005 = 2.0×10^{10}

Related Products:

- ig® 5-Alpha Chemically Comp. Cells (Cat.# 1031-12)
- ig® 10B Chemically Comp. Cells (Cat.# 1011-12)
- T4 DNA Ligase (Cat.# 3212)
- i7® High Fidelity DNA Polymerase (Cat.# 3254)
- igFusion™ Cloning Kit (Cat.# 4111)

Ordering Information:

- Order online within the USA. Place orders on **www.intactgenomics.com** using our secure Shopping Cart.
- Order by email, phone, or fax.
  Email: **sales@intactgenomics.com**
  Phone: (314) 942-3655 **|** Toll-free : 855-835-7172 **|** Fax: (314) 942-3656
- Order via our distributors.

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Our hours are Monday - Friday, 8AM to 5PM, U.S. Central Standard Time.

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