



# igMax™ DH10B ElectroCompetent Cells

## Manual

Catalog #	1284-24
Package Size	6x100µl



### Important!

#### **-80°C Storage Required**

- \* Immediately inspect packages
- \* Freeze upon receipt



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**Intact Genomics, Inc.**

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### Description:

igMax™ DH10B derivative electrocompetent cells are suitable for demanding cloning situations such as synthetic bio-applications, BAC cloning, assembling large multi-DNA fragments or cloning difficult targets requiring the greatest number of transformants possible. Utilizing proprietary manufacturing methods, these cells allow for effective transformation of all large DNA molecules ( $\geq 10\text{kb}$  up to  $350\text{kb}$ )!

### Specifications:

**Competent cell type:** Electrocompetent

**Derivative of:** DH10B

**Species:** *E. coli*

**Format:** Tubes

**Transformation efficiency:**  $\geq 5 \times 10^{10}$  cfu/ $\mu\text{g}$  pUC19 DNA

**Blue/white screening:** Yes

**Shipping condition:** Dry ice

### Components & Storage:

- igMax™ DH10B Electrocompetent cells:  $-80\text{ }^{\circ}\text{C}$
- pUC19 control DNA:  $-20\text{ }^{\circ}\text{C}$
- Recovery medium:  $4\text{ }^{\circ}\text{C}$

### Reagents Needed for One Reaction:

- igMax™ DH10B Electrocompetent cells:  $25\text{ }\mu\text{l}$
- DNA (or pUC19 Control,  $10\text{ pg}/\mu\text{l}$ ):  $1\text{ }\mu\text{l}$
- Recovery medium:  $1\text{ ml}$

## Genotype:

F - mcrA  $\Delta$ (mrr-hsdRMS-mcrBC) endA1 recA1  $\phi$ 80dlacZ $\Delta$ M15  $\Delta$ lacX74 araD139  $\Delta$ (ara, leu 7697 galU galK rpsL (StrR) nupG  $\lambda$

## Genomic Features:

igMax™ Electrocompetent DH10B cells have the following features:

- $\geq 5 \times 10^{10}$  cfu/ $\mu$ g efficiency with electroporation.
- $5 \sim 10 \times 10^7$  for 100~150 kb large DNA

## Quality Control:

Transformation efficiency is tested by using the pUC19 control DNA supplied with the kit and the high efficiency transformation protocol listed below. Transformation efficiency should be  $\geq 1 \times 10^{10}$  CFU/ $\mu$ g pUC19 DNA. Untransformed cells are tested for appropriate antibiotic sensitivity.

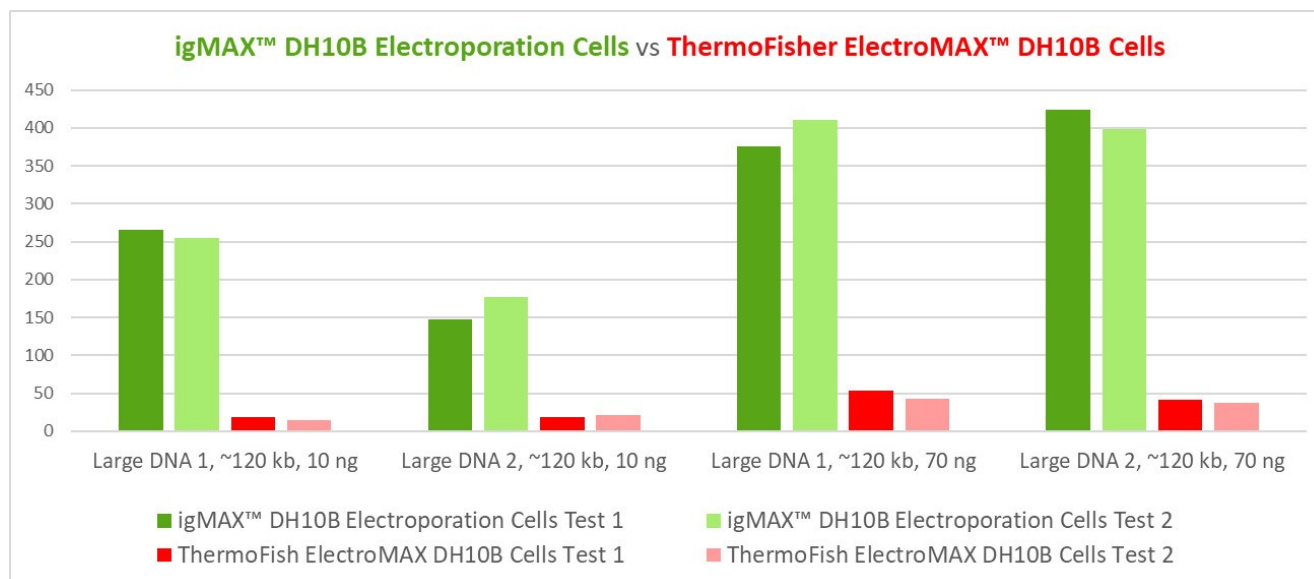
## General Guidelines:

Follow these guidelines when using igMax™ DH10B ElectroCompetent Cells:

- Handle competent cells gently as they are highly sensitive to changes in temperature or mechanical lysis caused by pipetting.
- Thaw competent cells on ice and transform cells immediately following thawing. After adding DNA, mix by tapping the tube gently. Do not mix cells by pipetting or vortexing.

**Note:** A high-voltage electroporation apparatus such as Bio-Rad Gene Pulser II #165-2105, capable of generating field strengths of 16 kV/cm is required.

## Transformation Protocol:



Extremely large DNA molecules can effectively be transformed at a rate much higher than ANY competitor, as shown in the figure above. Use this procedure to transform igMax™ DH10B Electrocompetent Cells. Do not use these cells for chemical transformation.

- 1) Place sterile cuvettes and microcentrifuge tubes on ice.
- 2) Remove competent cells from the -80 °C freezer and thaw completely on wet ice (10-15 minutes).
- 3) Aliquot 1 µl (1 pg-10 ng) of DNA to the chilled microcentrifuge tubes on ice.
- 4) When the cells are thawed, add 20 µl of cells to each DNA tube on ice and mix gently by tapping 4-5 times. For the pUC19 control, add 1 µl of (10 pg/µl) DNA to the 25 µl of cells on ice. Mix well by tapping. Do not pipette up and down or vortex to mix, this can harm cells and decrease transformation efficiency.
- 5) Pipette 26 µl of the cell/DNA mixture into a chilled electroporation cuvette without introducing bubbles. Quickly flick the cuvette downward with your wrist to deposit the cells across the bottom of the well and then electroporate.
- 6) Immediately add 974 µl of Recovery Medium or any other medium of choice to the cuvette, pipette up and down three times to re-suspend the cells. Transfer the cells and Recovery Medium to a culture tube.
- 7) Incubate tubes at 37 °C for 1 hour at 210 rpm.
- 8) Dilute the cells as appropriate then spread 20-200 µl cells onto a pre-warmed selective plate. For the pUC19 control, plate 50 µl of diluted transformants onto an LB plate containing 100 µg/ml ampicillin. Use sterilized spreader or autoclaved ColiRoller™ plating beads to spread evenly.
- 9) Incubate the plates overnight at 37 °C.

## Example Calculation of Transformation Efficiency:

Transformation Efficiency (TE) is defined as the number of colony forming units (cfu) produced by transforming 1µg of plasmid into a given volume of competent cells.

$$TE = \text{Colonies}/\mu\text{g}/\text{Dilution}$$

Transform 1 µl of (10 pg/µl) pUC19 control plasmid into 25 µl of cells, add 975 µl of Recovery Medium. Dilute 10 µl of this in 990 µl of Recovery Medium and plate 50 µl. Count the colonies on the plate the next day. If you count 250 colonies, the TE is calculated as follows:

$$\text{Colonies} = 250$$

$$\mu\text{g of DNA} = 0.00001$$

$$\text{Dilution} = 25/1000 \times 10/1000 = 0.00025$$

$$TE = 250/.00001/.00025 = 10.0 \times 10^{10}$$

## Related Products:

- DirectPlate™ 10B Chemically Competent Cells (Cat.# 1015-12 )
- DirectPlate™ DH5-Alpha Chemically Competent Cells (Cat.# 1013-12)
- DirectPlate XL Cells (Cat.# 1094-06)
- ig® XL1 Blue Max Chemically Competent Cells (Cat.# 1023-12 )
- Custom Competent Cells - Contact us for your next project!

## Ordering Information:

- Order online within the USA. Place orders on [www.intactgenomics.com](http://www.intactgenomics.com) using our secure Shopping Cart.
- Order by email, phone, or fax.  
Email: [sales@intactgenomics.com](mailto:sales@intactgenomics.com)  
Phone: (314) 942-3655 | Toll-free : 855-835-7172 | Fax: (314) 942-3656
- Order via our distributors.

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Our hours are Monday - Friday, 8AM to 5PM, U.S. Central Standard Time.

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