



ig® HB101 ElectroCompetent Cells

Manual

Catalog #	1271-12	1271-48
Package Size	6x50 µl	24x50 µl



Important!

-80°C Storage Required

- * Immediately inspect packages
- * Freeze upon receipt



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Intact Genomics, Inc.

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Description:

Intact Genomics (ig[®]) HB101 electrocompetent *E. coli* cells are suitable for high efficiency transformation in a wide variety of applications such as cloning and sub-cloning. *E. coli* HB101 is a K12 x B hybrid strain, containing the recA13 mutation that minimizes recombination and helps insert stability. In addition, it carries the hsdS20(rB-mB-) restriction minus genotype which prevents cleavage of cloned DNA by endogenous restriction enzymes. HB101 strain does not support Alpha-complementation for blue/white screening.

Specifications:

Competent cell type: Electrocompetent

Species: *E. coli*

Derivative of: HB101

Format: Tubes

Transformation efficiency: $\geq 4 \times 10^{10}$ cfu/ μ g pUC19 DNA

Blue/white screening: Yes

Shipping condition: Dry ice

Reagents Needed for One Reaction:

- ig[®] HB101 electrocompetent cells: 25 μ l
- DNA (or pUC19 Control, 10 pg/ μ l): 1 μ l
- Recovery medium: 1 ml

Storage:

- ig[®] HB101 electrocompetent cells: -80 °C
- pUC19 control DNA: -20 °C
- Recovery medium: 4 °C

Genotype:

F- Lambda- araC14 leuB6(Am) DE(gpt-proA)62 lacY1 glnX44(AS) galK2(Oc) recA13 rpsL20(strR) xylA5 mtl-1 thiE1 hsdS20(rB-, mB-)

Quality Control:

Transformation efficiency is tested by using the pUC19 control DNA supplied with the kit and the high efficiency transformation protocol listed below. Transformation efficiency should be $\geq 4 \times 10^{10}$ CFU/ μ g pUC19 DNA. Untransformed cells are tested for appropriate antibiotic sensitivity.

General Guidelines:

Follow these guidelines when using ig[®] HB101 Electrocompetent Cells:

- Handle competent cells gently as they are highly sensitive to changes in temperature or mechanical lysis caused by pipetting.
- Thaw competent cells on ice and transform cells immediately following thawing. After adding DNA, mix by tapping the tube gently. Do not mix cells by pipetting or vortexing.

Note: A high-voltage electroporation apparatus such as Bio-Rad Gene Pulser II #165-2105, capable of generating field strengths of 16 kV/cm is required.

Calculation of Transformation Efficiency:

Transformation Efficiency (TE) is defined as the number of colony forming units (cfu) produced by transforming 1 μ g of plasmid into a given volume of competent cells.

$$TE = \text{Colonies}/\mu\text{g}/\text{Dilution}$$

Transform 1 μ l of (10 pg/ μ l) pUC19 control plasmid into 25 μ l of cells, add 950 μ l of Recovery Medium. Dilute 10 μ l of this in 990 μ l of Recovery Medium and plate 50 μ l. Count the colonies on the plate the next day. If you count 100 colonies, the TE is calculated as follows:

$$\text{Colonies} = 100$$

$$\mu\text{g of DNA} = 0.00001$$

$$\text{Dilution} = 50/1000 \times 10/1000 = 0.0005$$

$$TE = 100/.00001/.0005 = 2.0 \times 10^{10}$$

Transformation Protocol:

Use this procedure to transform ig[®] HB101 electrocompetent cells. Do not use these cells for chemical transformation.

- 1) Place sterile cuvettes and microcentrifuge tubes on ice.
- 2) Remove competent cells from the -80 °C freezer and thaw completely on wet ice (10-15 minutes).
- 3) Aliquot 1 µl (1 pg-10 ng) of DNA to the chilled microcentrifuge tubes on ice.
- 4) When the cells are thawed, add 25 µl of cells to each DNA tube on ice and mix gently by tapping 4-5 times. For the pUC19 control, add 1 µl of (10 pg/µl) DNA to the 25 µl of cells on ice. Mix well by tapping. Do not pipette up and down or vortex to mix, this can harm cells and decrease transformation efficiency.
- 5) Pipette 26 µl of the cell/DNA mixture into a chilled electroporation cuvette without introducing bubbles. Quickly flick the cuvette downward with your wrist to deposit the cells across the bottom of the well and then electroporate.
- 6) Immediately add 974 µl of Recovery Medium or any other medium of choice to the cuvette, pipette up and down three times to re-suspend the cells. Transfer the cells and Recovery Medium to a culture tube.
- 7) Incubate tubes at 37 °C for 1 hour at 210 rpm.
- 8) Dilute the cells as appropriate then spread 20-200 µl cells onto a pre-warmed selective plate. For the pUC19 control, plate 50 µl of diluted transformants onto an LB plate containing 100 µg/ml ampicillin. Use sterilized spreader or autoclaved ColiRoller™ plating beads to spread evenly.
- 9) Incubate the plates overnight at 37 °C.

Related Products:

- ig[®] 5-Alpha Chemically Comp. Cells (Cat.# 1031-12)
- ig[®] 10B Chemically Comp. Cells (Cat.# 1011-12)
- T4 DNA Ligase (Cat.# 3212)
- i7[®] High Fidelity DNA Polymerase (Cat.# 3254)
- igFusion™ Cloning Kit (Cat.# 4111)

Ordering Information:

- Order online within the USA. Place orders on **www.intactgenomics.com** using our secure Shopping Cart.
- Order by email, phone, or fax.
Email: **sales@intactgenomics.com**
Phone: (314) 942-3655 | Toll-free : 855-835-7172 | Fax: (314) 942-3656
- Order via our distributors.

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Our hours are Monday - Friday, 8AM to 5PM, U.S. Central Standard Time.

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