



ig® 5-alpha Chemically Competent Cells

Manual

Catalog #	Package Size
1031-06	6x50 µl
1031-12	12x50 µl
1031-24	24x50 µl
1031-48	48x50 µl
1032-12	6x100 µl
1032-24	12x100 µl
1034-24	6x200 µl
1034-48	12x200 µl
1038-96	96 well x 20 µl



Important!

-80°C Storage Required

- * Immediately inspect packages
- * Freeze upon receipt



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Intact Genomics, Inc.

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Description:

Intact Genomics (ig[®]) 5-alpha chemically competent *E. coli* cells are suitable for high efficiency transformation in a wide variety of applications such as cloning and sub-cloning.

Specifications:

Competent cell type: Chemically Competent

Species: *E. coli*

Derivative of: DH5a[™]

Format: Tubes

Transformation efficiency: $\geq 1.0 \times 10^9$ cfu/ μ g pUC19 DNA

Blue/white screening: Yes

Shipping condition: Dry ice

Reagents Needed for One Reaction:

- ig[®] 5-alpha chemically competent cells: 50 μ l
- DNA (or pUC19 Control, 10 pg/ μ l): 1 μ l
- Recovery medium: 1 ml

Product Components and Storage:

- ig[®] 5-alpha competent cells: -80 °C
- pUC19 control DNA: -20 °C
- Recovery medium: 4 °C

Genotype:

Φ 80 Δ (lacZ)M15 fhuA2 Δ (argF-lacZ)U169 phoA glnV44 gyrA96 recA1 relA1 endA1 thi-1hsdR17

Quality Control:

Transformation efficiency is tested by using the pUC19 control DNA supplied with the kit and the high efficiency transformation protocol listed below. Transformation efficiency should be $\geq 1 \times 10^9$ CFU/ μ g pUC19 DNA. Untransformed cells are tested for appropriate antibiotic sensitivity.

General Guidelines:

Follow these guidelines when using ig[®] 5-alpha chemically competent *E. coli*.

- Handle competent cells gently as they are highly sensitive to changes in temperature or mechanical lysis caused by pipetting.
- Thaw competent cells on ice, and transform cells immediately following thawing. After adding DNA, mix by tapping the tube gently. Do not mix cells by pipetting or vortexing.

Calculation of Transformation Efficiency:

Transformation Efficiency (TE) is defined as the number of colony forming units (cfu) produced by transforming 1 μ g of plasmid into a given volume of competent cells.

$$TE = \text{Colonies}/\mu\text{g}/\text{Dilution}$$

Transform 1 μ l of (10 pg/ μ l) pUC19 control plasmid into 50 μ l of cells, add 950 μ l of Recovery Medium. Dilute 10 μ l of this in 990 μ l of Recovery Medium and plate 50 μ l. Count the colonies on the plate the next day. If you count 100 colonies, the TE is calculated as follows:

$$\text{Colonies} = 100$$

$$\mu\text{g of DNA} = 0.00001$$

$$\text{Dilution} = 50/1000 \times 10/1000 = 0.0005$$

$$TE = 100/.00001/.0005 = 2.0 \times 10^{10}$$

High Efficiency Transformation Protocol:

Use this procedure to transform ig[®] 5-alpha chemically competent cells. We recommend verifying the transformation efficiency of the cells using the pUC19 control DNA supplied with the kit. Do not use these cells for electroporation.

- 1) Remove competent cells from the -80 °C freezer and thaw completely on wet ice (10-15 minutes).
- 2) Aliquot 1-5 µl (1 pg-100 ng) of DNA to the chilled microcentrifuge tubes on ice.
- 3) When the cells are thawed, add 50 µl of cells to each DNA tube on ice and mix gently by tapping 4- 5 times. For the pUC19 control, add 1 µl of (10 pg/ µl) DNA to a chilled microcentrifuge tube, prior to adding 50 µl of cells. Mix well by tapping. Do not pipette up and down or vortex to mix, this can harm cells and decrease transformation efficiency.
- 4) Incubate the cells with DNA on ice for 30 minutes.
- 5) After 30 minute ice incubation, heat shock the cells at 42 °C for 45 seconds.
- 6) Transfer the tubes to ice for 2 minutes.
- 7) Add 950 µl of Recovery Medium or any other medium of choice to each tube.
- 8) Incubate tubes at 37 °C for 1 hour at 210 rpm.
- 9) Spread 50 µl to 200 µl from each transformation on Pre-warmed selection plates. We recommend plating two different volumes to ensure that at least one plate will have well-spaced colonies. For the pUC19 control, plate 50 µl on an LB plate containing 100 µg/ ml ampicillin. Use sterilized spreader or autoclaved ColiRoller™ plating beads to spread evenly.
- 10) Incubate the plates overnight at 37 °C.

5 Minute Transformation Protocol:

The following procedure results in only ~10% of the transformation efficiency as the protocol listed above.

- 1) Remove competent cells from the -80 °C freezer and thaw in your hand.
- 2) Aliquot 1-5 µl (1 pg-100 ng) of DNA to the microcentrifuge tubes. Do not pipette up and down or vortex to mix, this can harm cells and decrease transformation efficiency.
- 3) Incubate the cells with DNA on ice for 2 minutes.
- 4) After 2 minute ice incubation, heat shock the cells at 42 °C for 45 seconds.
- 5) Transfer the tubes to ice for 2 minutes.
- 6) Add 950 µl of Recovery Medium at room temperature or any other medium of choice to each tube. Immediately spread 50 µl to 200 µl from each transformation on prewarmed selection plates. We recommend plating two different volumes to ensure that at least one plate will have well-spaced colonies. For the pUC19 control, plate 50 µl on an LB plate containing 100 µg/ml ampicillin. Use sterilized spreader or autoclaved ColiRoller™ plating beads to spread evenly.
- 7) Incubate the plates overnight at 37 °C.

Related Products:

- ig[®] 10B Chemically Comp. Cells (Cat.# 1011-12)
- ig[®] 5-alpha Electroporation Comp. Cells (Cat.# 1232-12)
- T4 DNA Ligase (Cat.# 3212)
- i7[®] High Fidelity DNA Polymerase (Cat.# 3254)

Ordering Information:

- Order online within the USA. Place orders on www.intactgenomics.com using our secure Shopping Cart.
- Order by email, phone, or fax.
Email: sales@intactgenomics.com
Phone: (314) 942-3655 | Toll-free : 855-835-7172 | Fax: (314) 942-3656
- Order via our distributors.

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Technical Support & Customer Services

Intact Genomics (IG®) is dedicated to customer satisfaction regarding the use of our products for your research needs. Each new lot of our products is thoroughly tested to ensure it meets high quality standards and provides excellent results. We appreciate your business and your feedback regarding the performance of our products in your applications. Please follow the instructions carefully and contact us if additional assistance is needed.

Our hours are Monday - Friday, 8AM to 5PM, U.S. Central Standard Time.

Intact Genomics, Inc.

11840 Westline Industrial Drive, Suite 120,
St. Louis, MO. 63146, USA

Phone: (314) 942-3655 | **Toll-free :** 855-835-7172 | **Fax:** (314) 942-3656

Email: sales@intactgenomics.com | ig@intactgenomics.com

Website: www.intactgenomics.com

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