



Staphylococcus aureus (S. aureus) RN4220 **ElectroCompetent Cells**

Manual

Catalog #	1294-40
Package Size	5x200µl



Important!

-80°C Storage Required

- * Immediately inspect packages
- * Freeze upon receipt



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Intact Genomics, Inc.

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Description:

Intact Genomics (ig®) *Staphylococcus aureus* (*S. aureus*) RN4220 electrocompetent cells are optimized to provide high transformation efficiencies making them ideal for various applications. This strain is commonly used in studies involving virulence, resistance, metabolism and more. *Staphylococcus aureus* RN4220 is characterized by a mutation in the *sau1 hsdR* genes, making it restriction deficient and hence an excellent intermediate cloning host.

Specifications:

Competent cell type: Electrocompetent

Species: *S. aureus*

Strain: RN4220

Format: Tubes

Transformation efficiency: $\geq 1.0 \times 10^5$ cfu/ μ g

Blue/white screening: No

Shipping condition: Dry ice

Reagents Included:

- ig® *Staphylococcus aureus* (*S. aureus*) RN4220 Electrocompetent Cells
- Recovery medium

Storage:

- ig® *Staphylococcus aureus* (*S. aureus*) RN4220 Electrocompetent Cells: -80 °C
- Recovery medium: 4 °C

Quality Control:

Transformation efficiency is tested by using the control DNA supplied with the kit and using the high-efficiency transformation protocol. Transformation efficiency should be $\geq 1 \times 10^5$ CFU/ μ g DNA. Untransformed cells are tested for appropriate antibiotic sensitivity.

General Guidelines:

Follow these guidelines when using ig® *S. aureus* Electrocompetent cells:

- Handle competent cells gently as they are highly sensitive to changes in temperature or mechanical lysis caused by pipetting.
- Thaw competent cells on ice and transform cells immediately following thawing. After adding DNA, mix by tapping the tube gently. Do not mix cells by pipetting or vortexing.

Biosafety Level: 2

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the following publication: U.S. Department of Health and Human Services, Public Health Service, Centers for Disease Control and Prevention, and National Institutes of Health. Biosafety in Microbiological and Biomedical Laboratories. 5th ed. Washington, DC: U.S. Government Printing Office, 2009; see:

www.cdc.gov/biosafety/publications/bmbl5/index.htm.

BSL2 facility is required for purchase and use of this product

Calculation of Transformation Efficiency:

Transformation Efficiency (TE) is defined as the number of colony forming units (cfu) produced by transforming 1µg of plasmid into a given volume of competent cells.

$$TE = \text{Colonies}/\mu\text{g}/\text{Plated}$$

Transform 1 µl of (500 ng/µl) of the control plasmid into 25 µl of cells, add 500 µl of Recovery Medium. Recover for one hours and plate 100 µl. Count the colonies on the plate in two days. If you count 1000 colonies, the TE is calculated as follows:

$$\text{Colonies} = 1,000$$

$$\mu\text{g of DNA} = 0.05$$

$$\text{Dilution} = 100/500 = 0.2$$

$$TE = 1000/.05/.2 = 1 \times 10^5$$

Transformation Protocol:

Use this procedure to transform ig[®] *Staphylococcus aureus* ElectroCompetent Cells. Do not use these cells for chemical transformation.

- 1) Place sterile cuvettes and microcentrifuge tubes on ice.
- 2) Remove competent cells from the -80 °C freezer and thaw completely on wet ice (10-15 minutes).
- 3) Aliquot 1 µl (10pg -1 µg) of DNA to the chilled microcentrifuge tubes on ice.
- 4) When the cells are thawed, add 25 µl of cells to each DNA tube on ice and mix gently by tapping 4-5 times. For the control, add 1 µl of (50 ng/µl) DNA to the 25 µl of cells on ice. Mix well by tapping. Do not pipette up and down or vortex to mix, this can harm cells and decrease transformation efficiency.
- 5) Pipette 26 µl of the cell/DNA mixture into a chilled electroporation cuvette without introducing bubbles. Quickly flick the cuvette downward with your wrist to deposit the cells across the bottom of the well and then electroporate.
- 6) Immediately add 500µl of Recovery Medium or any other medium of choice to the cuvette, pipette up and down three times to re-suspend the cells. Transfer the cells and Recovery Medium to an Eppendorf tube.
- 7) Incubate tubes at 37 °C for 1 hours at 200 RPM.
- 8) Dilute the cells as appropriate then spread 20-200 µl cells onto a pre-warmed selective plate. For the control plasmid , you may plate 100 µl of undiluted transformation mix onto a YT plate containing chloramphenicol (12.5µg/µl). Use sterilized spreader or autoclaved ColiRoller[™] plating beads to spread evenly.
- 9) Incubate the plates for overnight at 37 °C.

Electroporation Settings:

Mode: Exponential protocol

Voltage (V): 1,800 V

Capacitance: 25 uFD

Resistance: 200 Ohms

Cuvette: 1 mm

Related Products:

- T4 DNA Ligase (Cat.# 3212)
- igFusion Cloning Kit (Cat.# 4111)
- i7® High Fidelity DNA Polymerase (Cat.# 3254)
- *E. coli* and other competent cells

Ordering Information:

- Order online within the USA. Place orders on **www.intactgenomics.com** using our secure Shopping Cart.
- Order by email, phone, or fax.
Email: **sales@intactgenomics.com**
Phone: (314) 942-3655 | Toll-free : 855-835-7172 | Fax: (314) 942-3656
- Order via our distributors.

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Our hours are Monday - Friday, 8AM to 5PM, U.S. central standard time.

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