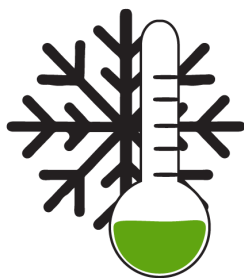


# ER2738 Phage Display Electrocompetent Cells

Manual



## Important!

### **-80°C Storage Required**

- \* Immediately inspect packages
- \* Freeze upon receipt

**FOR RESEARCH USE ONLY. NOT FOR HUMAN OR DIAGNOSTIC USE**

**Catalog #1217-12 and 1217-24**

**Intact Genomics, Inc.**

[www.intactgenomics.com](http://www.intactgenomics.com)



## Description:

Intact Genomics (ig<sup>®</sup>) ER2738 Phage Display ElectroCompetent Cells are suitable for protein expression and preparation of antibody or peptide phage display libraries. ER2738 cells are also useful for protein expression, M13 phage work, general cloning, and blue/ white screening.

## Reagents Included:

- ig<sup>®</sup> ER2738 phage display electrocompetent cells
- pUC19 Control
- Recovery medium

## Product Storage:

ig<sup>®</sup> ER2738 phage display electrocompetent cells: -80 °C

pUC19 control DNA: -20 °C

Recovery medium: 4 °C

## Genomic Features:

Intact Genomics ER2738 phage display electrocompetent cells have the following features:

- $>4 \times 10^{10}$  cfu/ $\mu$ g efficiency with electroporation.
- Amber suppressor strain (glnV)

## Genotype:

*[F'proA+B+ lacIq Δ(lacZ)M15 zcf::Tn10 (tetr)] fhuA2 glnVΔ(lac-proAB) thi-1Δ(hsdS-mcrB)5*

## Quality Control:

Transformation efficiency is tested by using the pUC19 control DNA supplied with the kit and using the protocol given below. Transformation efficiency should be  $>4 \times 10^{10}$  CFU/ $\mu$ g pUC19 DNA. Untransformed cells are tested for appropriate antibiotic sensitivity.

## Calculation of Transformation Efficiency:

Transformation Efficiency (TE) is defined as the number of colony forming units (cfu) produced by transforming 1 $\mu$ g of plasmid into a given volume of competent cells. TE = Colonies/ $\mu$ g/Dilution

Transform 1  $\mu$ l of (10 pg/ $\mu$ l) pUC19 control plasmid into 50  $\mu$ l of cells, add 950  $\mu$ l of Recovery Medium. Dilute 10  $\mu$ l of this in 990  $\mu$ l of Recovery Medium and plate 50  $\mu$ l. Count the colonies on the plate the next day. If you count 200 colonies, the TE is calculated as follows:

Colonies = 200

$\mu$ g of DNA = 0.00001

Dilution =  $50/1000 \times 10/1000 = 0.0005$

TE =  $200/.00001/.0005 = 4.0 \times 10^{10}$

## **General Guidelines:**

Follow these guidelines when using Intact Genomics ER2738 phage display electrocompetent cells:

Handle competent cells gently as they are highly sensitive to changes in temperature or mechanical lysis caused by pipetting.

Thaw competent cells on ice, and transform cells immediately following thawing. After adding DNA, mix by tapping the tube gently. Do not mix cells by pipetting or vortexing.

**Note:** A high-voltage electroporation apparatus such as Bio-Rad Gene Pulser II #165-2105, capable of generating field strengths of 16 kV/cm is required.

## **Transformation Protocol:**

Use this procedure to transform Intact Genomics ER2738 phage display electrocompetent cells. Do not use these cells for chemically transformation.

- 1) Place sterile cuvettes and microcentrifuge tubes on ice.
- 2) Remove competent cells from the -80 °C freezer and thaw completely on wet ice (10-15 minutes).
- 3) Aliquot 1 µl (1 pg-10 ng) of DNA to the chilled microcentrifuge tubes on ice.
- 4) When the cells are thawed, add 25 µl of cells to each DNA tube on ice and mix gently by tapping 4-5 times. For the pUC19 control, add 1 µl of (10 pg/µl) DNA to the 25 µl of cells on ice. Mix well by tapping. Do not pipette up and down or vortex to mix, this can harm cells and decrease transformation efficiency.
- 5) Pipette 26 µl of the cell/DNA mixture into a chilled electroporation cuvette without introducing bubbles. Quickly flick the cuvette downward with your wrist to deposit the cells across the bottom of the well and then electroporate.
- 6) Immediately add 974 µl of Recovery Medium or any other medium of choice to the cuvette, pipette up and down three times to re-suspend the cells. Transfer the cells and Recovery Medium to a culture tube.
- 7) Incubate tubes at 37 °C for 1 hour at 210 rpm.
- 8) Dilute the cells as appropriate then spread 20-200 µl cells onto a pre-warmed selective plate. For the pUC19 control, plate 50 µl of diluted transformants onto an LB plate containing 100 µg/ml ampicillin. Use sterilized spreader or autoclaved ColiRoller™ plating beads to spread evenly.
- 9) Incubate the plates overnight at 37 °C.

## Related Products

- TG1 Phage Display Electrocompetent Cells (Cat#1264-24)
- SS320 Phage Display Electrocompetent Cells (Cat#1264-24)
- Taq DNA Polymerase 2x Premix (Cat.# 3249)
- T4 DNA Ligase (Cat.# 3212)
- ig<sup>®</sup> 10B Chemically Competent Cells (Cat.# 1011-12)

## Technical Support

Intact Genomics (IG<sup>®</sup>) is dedicated to customer satisfaction regarding the use of our products for your research needs. We test our products thoroughly to ensure they conform to the highest quality standards and provide excellent results when following the protocol's specifications. Please follow the protocol information provided in this manual carefully and contact our customer and technical support team with any questions or comments you may have regarding this or our other products.

### IG Technical Support

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